

HETEROCYCLES, Vol. 65, No. 8, 2005, pp. 1967 - 1973

Received, 15th April, 2005, Accepted, 27th May, 2005, Published online, 27th May, 2005

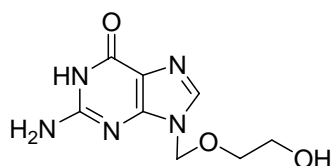
A PRACTICAL SYNTHESIS OF 8-HYDROXYACYCLOVIR AND 9-(CARBOXYMETHOXYMETHYL)GUANINE, METABOLITES OF ACYCLOVIR

Teruo Kutsuma,^a Yasuo Sakai,^b Hiroyuki Ouchi,^b Noriaki Shiina,^c Takehiro Yamagishi,^a Shiroshi Shibuya,^a and Tsutomu Yokomatsu^{a,*}

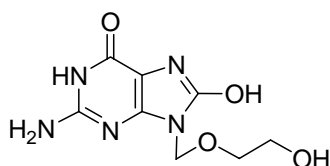
^aSchool of Pharmacy, Tokyo University of Pharmacy & Life Science, 1432-1 Horinouchi, Hachioji, Tokyo 192-0392, Japan. ^bToa Pharmaceuticals Co., Ltd., 26 Sango, Toyama 939-3548, Japan. ^cJunsei Chemical Co., Ltd., 1-6 Ohmano, Koshigaya, Saitama 343-0844, Japan. E-mail: yokomatu@ps.toyaku.ac.jp

Abstract – Improved methods for the synthesis of 8-hydroxyacyclovir and 9-(carboxymethoxymethyl)guanine, metabolites of acyclovir, were examined. The methods were found to be useful for practical preparation of 8-hydroxyacyclovir and 9-(carboxymethoxymethyl)guanine of high purity. Careful spectroscopic analysis of 8-hydroxyacyclovir in DMSO-*d*₆ suggested that it may exist in an 8-oxo tautomer rather than an 8-hydroxy tautomer under the solution conditions.

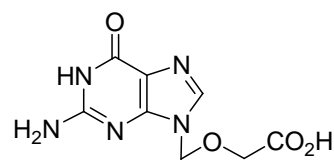
9-(2-Hydroxyethoxymethyl)guanine (**1**) (acyclovir), the acyclic nucleoside analogue of guanosine, is a selective inhibitor of the replication of herpes simplex virus (HSV) types 1 and 2 and varicella-zoster virus.¹ Many synthetic and metabolic studies of acyclovir have been reported in the last decade.² The metabolic disposition of acyclovir has been investigated in humans and several experimental species including mice, rats and dogs. In these metabolic studies, 8-hydroxy-9-(2-hydroxyethoxymethyl)guanine (8-hydroxyacyclovir) (**2**) and 9-(carboxymethoxymethyl)guanine (CMMG) (**3**) have been identified as major metabolites for humans and animals.³



acyclovir (**1**)



8-hydroxy acyclovir (**2**)



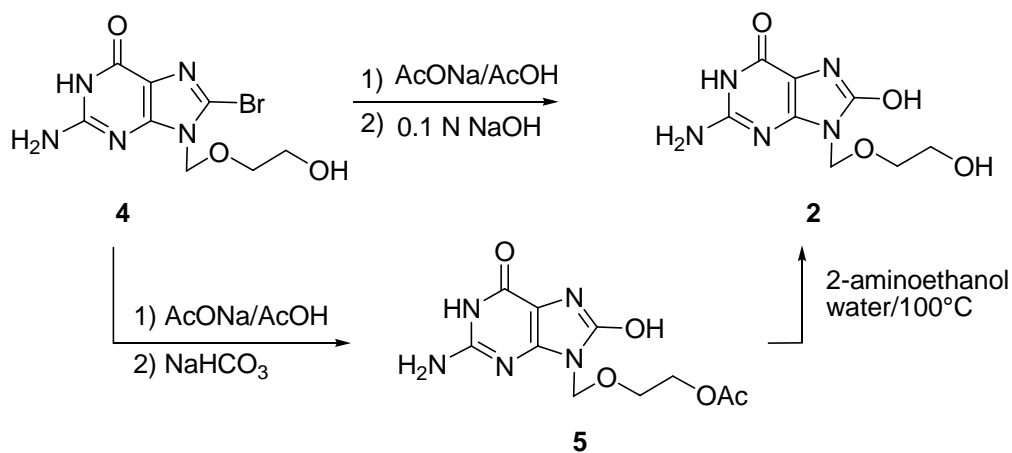
CMMG (**3**)

In the course of our investigation directed toward developing novel prodrugs of acyclovir, we required a large quantity of 8-hydroxyacyclovir and **CMMG** of high purity for the metabolic studies. Although a method for the synthesis of 8-hydroxyacyclovir (**2**) has been reported in the literature,⁴ in our experience, the method could not be applied to large-scale synthesis. The structural analysis of **2** was not studied in detail. Moreover, synthetic methods for **CMMG** were not disclosed in the literature. In this paper we wish to describe experimental details for a practical synthesis of 8-hydroxyacyclovir and **CMMG** with their full physical constants.

SYNTHESIS OF 4-HYDROXYACYCLOVIR

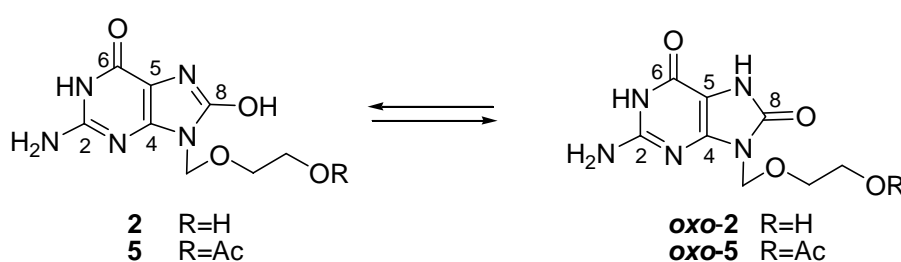
Robins *et al.* reported that 8-hydroxyacyclovir (**2**) could be prepared from 8-bromoacyclovir (**4**) by treatment with sodium acetate in glacial acetic acid, followed by neutralization with 0.1N NaOH.⁴ However, our attempted application of the reported procedure for the synthesis in large scale was not successful, due to production of a mixture of **2** and the corresponding acetates in varying ratios, which were not readily separated. Then, we examined a modified procedure for obtaining **2** in large scale (Scheme 1).

Treatment of 8-bromoacyclovir (**4**) with sodium acetate in glacial acetic acid at reflux for 6 h, followed by neutralization with powdered NaHCO₃ gave the acetate (**5**) in 75% yield. The results suggest that the initial-formed 8-acetoxyacyclovir is transformed to **5** *via* concomitant migration of the acetyl group to the primary hydroxyl group under the conditions. In this procedure, we readily obtained multi-gram quantities of **5** of high purity after single recrystallization of the crude materials. When **5** was treated with 2-aminoethanol in boiling water for 5 h,⁶ hydrolysis of the acetate group cleanly occurred to give the requisite 8-hydroxyacyclovir (**2**), which could be isolated in 96% yield after single recrystallization of the crude materials from 25% aqueous acetic acid. By using the step-wise method *via* the acetate (**5**), we obtained several grams of extremely pure 8-hydroxyacyclovir (**2**) in 72% yield for two steps.



Scheme 1

Although compound (**2**) was believed for a long time albeit without substantiating evidence to be in the 8-hydroxy tautomer, theoretically, **2** would exist in an equilibrium between the 8-hydroxy tautomer and 8-oxo tautomer (*oxo-2*) (Scheme 2). Structural analysis of 8-hydroxy-2'-deoxyguanosine (**8-OH-dG**) has been reported by Culp and co-worker.⁷ They found that chemical shifts of 5-C for **8-OH-dG** in solution of pH 6.0–14 are markedly dependent upon the pH value. The 5-C resonance exhibited a significant downfield chemical shift of *ca.* 10 ppm from 101 ppm with increasing pH, while the remaining base carbons exhibit minimal effects in this pH range. These marked shift in 5-C indicates that increasing basicity of the solution increases the 8-enolate form of **8-OH-dG** in an equilibrium. Then we carefully analyzed the ¹³C NMR spectrum of **2** to elucidate the tautomeric structures in comparison with that of acyclovir and **CMMG** (Table 1). In the ¹³C NMR spectrum (DMSO-*d*₆) of **2**, 5-C and 6-C resonated at δ 98.6 and 153.7, respectively. The corresponding carbons for acyclovir (**1**) resonated at δ 116.5 and 156.9, respectively. Although no significant differences in the chemical shifts for 6-C were observed between **2** and acyclovir, the signal due to 5-C of **2** is significantly shifted upfield by *ca.* 18 ppm as compared with that of acyclovir. The 5-C of **2** resonates at approximately the same chemical shift to that of **8-OH-dG** in a solution of pH 6.0.⁷ While precise tautomeric ratios for **2** and *oxo-2* remain unclear at this stage, the results delineated in this section strongly suggest that 8-oxo tautomer (*oxo-2*) would be a presumed major tautomer for compound (**2**) under the solution conditions. In a similar manner, compound (**5**) was also estimated to exist as 8-oxo tautomer (*oxo-5*). A clear understanding of the pH-dependent tautomeric equilibrium for compounds (**2** and **5**) must await further experimentation.⁷



Scheme 2

Table 1. Selected ¹³C NMR spectral data of acyclovir derivatives^a

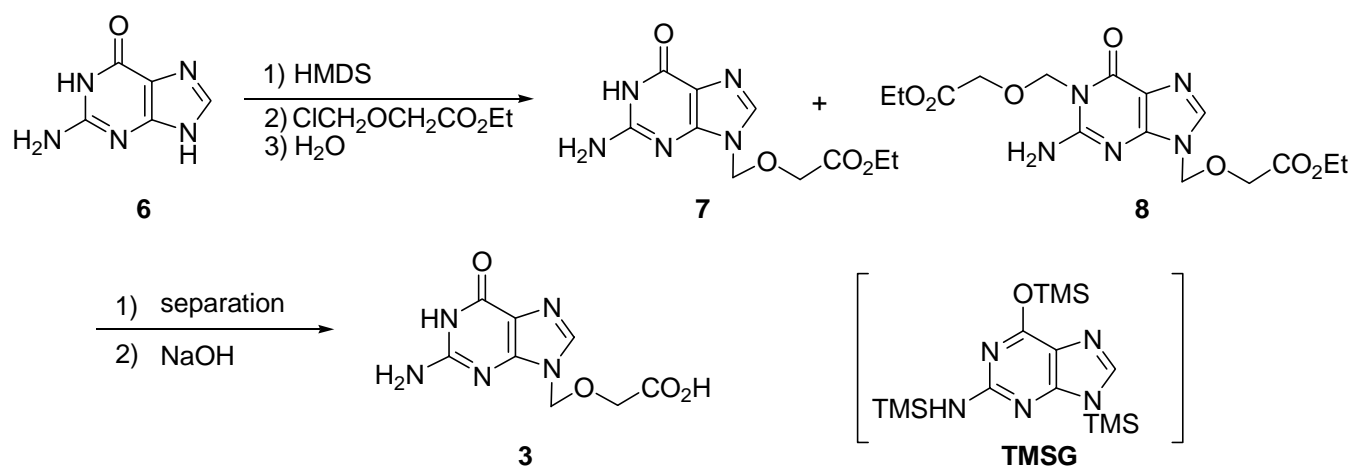
compound	chemical shifts, ppm				
	2-C	4-C	5-C	6-C	8-C
acyclovir (1)	153.9	151.5	116.5	156.9	137.9
CMMG	151.1	148.6	113.7	153.9	134.9
8-OH-acyclovir (2)	152.4	147.8	98.6	153.7	151.3
8-OH-acyclovir acetate (5)	151.0	147.4	98.5	152.1	153.4

^a All spectra were obtained in DMSO-*d*₆

SYNTHESIS OF 9-(CABOXYMETHOXYMETHYL)GUANINE (CMMG)

Although **CMMG** was proved to be a major metabolite of acyclovir by Schaeffer *et al.*,⁵ Wellcome Research Laboratories, to the best of our knowledge, details of synthetic methods for **CMMG** are not yet disclosed in scientific literature. We, therefore, examined a synthetic route for **CMMG** via *N*-glycosylation of guanine with ethyl (chloromethoxy)acetate (Scheme 3). The *N*-glycosylation of silylated guanine (**TMSG**) with ethyl (chloromethoxy)acetate has been reported by Russian chemists to give ethyl ester (**7**) of **CMMG**.⁸ However, in our experimentation, the reported procedure was very difficult to isolate **7** of high purity owing to production of a large quantity of inseparable gelatinous products. Then we examined modified conditions for *N*-glycosylation of **TMSG** with ethyl (chloromethoxy)acetate.

Although the Russian chemists used dichloroethane as a solvent for the *N*-glycosylation reaction, we examined several low-boiling solvents rather than dichloroethane to minimize formation of the gelatinous products. We found the gelatinous compounds were minimally formed when **TMSG**, prepared from guanine and hexamethyldisilazane (HMDS) in toluene, was treated with ethyl (chloromethoxy)acetate in toluene at 70°C for 12 h. The crude mixture was chromatographed on silica gel to give the desired **7** in 14.2% yield. We also isolated 1,9-bis(ethoxycarbonylmethoxymethyl)guanine (**8**) in 12.4% yield as a gelatinous product. Although the yield of **7** was modest, the method was available for synthesis of several grams quantities of **7**. Finally, compound (**7**) was hydrolyzed with aqueous NaOH to give the requisite **CMMG** (**3**) in quantitative yield.



Scheme 3

EXPERIMENTAL

All melting points are uncorrected. The NMR spectra were measured using 300 or 400 MHz spectrometers with DMSO-*d*₆ as the solvent and SiMe₄ as the internal standard. The assignment of ¹³C

carbon signals is based on DEPT data. IR spectra were recorded as a KBr pellet. Anhydrous toluene was purchased from Kanto Chemical Co., Ltd. and used without further purification. Mass spectra were recorded using electrospray ionization (ESI) techniques.

2-Amino-8-bromo-9-[(2-hydroxyethoxy)methyl]-1,9-dihydro-6H-purin-6-one (4)

To a stirred suspension of bromine (10.55 g, 66 mmol) in water (300 mL) was added a solution of acyclovir (13.52 g, 60 mmol) in water (2.3 L) at 40–45°C. The mixture was stirred at rt for 30 min and cooled with ice-cold water. The precipitates were filtered by suction and recrystallized from water to give **4** (15.9 g, 87%) as pale-yellow crystals: mp >300°C (lit.,⁴ mp ~280°C). ¹H NMR (300 MHz, DMSO-*d*₆) δ 3.42–3.51 (m, 4H, 2xCH₂), 4.68 (t, *J*=5.1 Hz, 1H, OH), 5.29 (s, 2H, NCH₂O), 6.62 (s, 2H, NH₂), 10.73 (s, 1H, NH). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 155.5 (6-C), 154.2 (2-C), 152.9 (4-C), 120.9 (8-C), 116.6 (5-C), 72.4 (N-CH₂-O), 70.8 (OCH₂CH₂OH), 59.9 (OCH₂CH₂OH). MS (ESI) *m/z* 304 (MH⁺). HRMS (ESI) calcd for C₈H₁₁N₅O₃Br (MH⁺): 304.0045. Found: 304.0045.

2-[(2-Amino-6,8-dioxo-1,6,7,8-tetrahydro-9H-purin-9-yl)methoxy]ethyl acetate (5)

A mixture of **4** (12.16 g, 40 mmol) and AcONa (24.61 g, 300 mmol) in AcOH (650 mL) was stirred under reflux for 6 h. The volatile components of the mixture were removed *in vacuo*. The resulting white solids were dissolved in hot water and neutralized by powdered NaHCO₃. The solution was cooled by ice water and the resulting precipitates were collected and recrystallized from water to give **5** (7.53 g, 78%) as colorless crystals: mp 268–269°C (decomp). ¹H NMR (300 MHz, DMSO-*d*₆) δ 1.96 (s, 3H, CH₃CO), 3.69 (t with small splits, *J*=4.7 Hz, 2H, CH₂), 4.06 (t with small splits, *J*=4.7 Hz, 2H, CH₂), 5.00 (s, 2H, NCH₂O), 6.50 (s, 2H, NH₂), 10.66 (s, 1H, NH), 10.69 (s, 1H, NH). ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 170.1 (C=O), 153.4 (8-C), 152.1 (6-C), 151.0 (2-C), 147.4 (4-C), 98.5 (5-C), 68.6 (NCH₂O), 66.6 (O-CH₂CH₂), 62.9 (CH₂CH₂O), 20.6 (CH₃). IR (KBr) 1723, 1689, 1647, 1602 cm⁻¹. MS (ESI) *m/z* 284 (MH⁺). Anal. Calcd for C₁₀H₁₃N₅O₅•H₂O: C, 39.86; H, 5.02; N, 23.25. Found: C, 39.88; H, 5.22; N, 23.47.

2-Amino-9-[(2-hydroxyethoxy)methyl]-7,9-dihydro-1H-purine-6,8-dione (8-hydroxyacyclovir) (2)

A suspension of **5** (5.5 g, 19.4 mmol) and 2-aminoethanol (2.33 g, 36.6 mmol) in water (85 mL) was heated under reflux for 5 h. The mixture was cooled. The resulting precipitates were collected and recrystallized from 25% aqueous acetic acid (16 mL) to give **2** (4.45 g, 95.5%) as colorless flakes: mp 289–290°C (decomp) (lit.,⁴ mp 260°C (decomp)). ¹H NMR (300 MHz, DMSO-*d*₆) δ 3.43–3.50 (m, 4H, 2xCH₂), 4.61 (t, *J*=5.7 Hz, 1H, OH), 4.99 (s, 2H, NCH₂O), 6.49 (s, 2H, NH₂), 10.63 (s, 1H, NH), 10.68 (s, 1H, NH). ¹³C NMR (75.5 MHz, DMSO-*d*₆) 153.7 (6-C), 152.4 (2-C), 151.3 (8-C), 147.8 (4-C), 98.6 (5-C),

70.8 (N-CH₂-O), 68.9 (O-CH₂-CH₂), 60.1 (CH₂-OH). IR (KBr) 1727, 1704, 1645, 1598 cm⁻¹. MS (ESI) *m/z* 242 (MH⁺). HRMS (ESI) calcd for C₈H₁₂N₅O₄ (MH⁺): 242.0889. Found: 242.0901. Anal. Calcd for C₈H₁₁N₅O₄•3/4H₂O: C, 37.72; H, 4.95; N, 27.50. Found: C, 37.97; H, 5.10; N, 27.56.

Ethyl (chloromethoxy)acetate

To a stirred suspension of ethyl glycolate (26 g, 250 mmol) and paraformaldehyde (6.76 g, 325 mmol) in toluene (700 mL) was passed a stream of dry hydrogen chloride for 15 min at -10°C. Then the mixture was treated with Na₂SO₄ (50 g) and stirred at the same temperature for 12 h. The temperature was elevated to 0°C. After being stirred for an additional 12 h, the solids were filtered. The volatile components of the filtrates were removed *in vacuo* at below 30°C to leave the title compound (23.5 g, 61.6%) as a colorless liquid. ¹H NMR (300 MHz, CDCl₃) δ 1.29 (t, *J*=7.2 Hz, 3H, CH₃), 4.20 (q, *J*=7.2 Hz, 2H, OCH₂CH₃), 4.28 (s, 2H, OCH₂CO-), 5.54 (s, 2H, ClCH₂O). This liquid was used for the next reaction without further purification.

Ethyl (2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)acetate (7) and Ethyl ({2-amino-1-[(2-ethoxy-2-oxoethoxy)methyl]-6-oxo-1,6-dihydro-9H-purin-9-yl}methoxy)acetate (8).

A mixture of guanine (6) (12.86 g, 85 mmol), hexamethyldisilazane (116.5 mL, 550 mmol), and (NH₄)₂SO₄ (1.0 g) in toluene (120 mL) was refluxed for 2 days. The mixture was cooled to rt. The resulting precipitates were filtered. The volatile components of filtrates were removed *in vacuo* to leave a yellow oil, which was treated with ethyl (chloromethoxy)acetate (13 g, 85 mmol) in toluene (120 mL) at 70°C for 20 h. The mixture was neutralized by Et₃N (8.7 g, 86 mmol) and the solvent was removed *in vacuo*. The resulting yellow solid was heated in boiling water (60 mL) for 1 h. After being cooled to rt, the solid materials (21.3 g) were collected and treated with a mixture of EtOH (50 mL) and CHCl₃ (200 mL) at reflux. The solid materials were removed by centrifugal separation. The supernatant was concentrated and the residue was chromatographed on silica gel. Elution with CHCl₃/EtOH(4:1) gave 8 (4.03 g, 12.4%) as white crystals: mp 191–193°C (from 70%EtOH). ¹H NMR (300 MHz, DMSO-*d*₆) δ 1.12–1.19 (m, 6H, 2xCH₃), 4.02–4.13 (m, 4H, 2xOCH₂CH₃), 4.26 (s, 2H, OCH₂CO), 4.28 (s, 2H, OCH₂CO), 5.54 (s, 2H, NCH₂O), 5.71 (s, 2H, NCH₂O), 7.15 (s, 2H, NH₂), 8.10 (s, 1H, 8-H). Anal. Calcd for C₁₅H₂₁N₅O₇: C, 46.99; H, 5.52; N, 18.27. Found: C, 46.83; H, 5.59; N, 17.70. Successive elution with CHCl₃/EtOH (7:3) gave white solids (3.92 g), which was recrystallized from water to give 7 (3.45 g, 14.2%) as colorless crystals: mp 223–224°C (lit.,⁸ mp 211–213°C). ¹H NMR (300 MHz, DMSO-*d*₆) δ 1.14 (t, *J*=7.2 Hz, 3H, CH₃), 4.05 (q, *J*=7.2 Hz, 2H, OCH₂CH₃), 4.19 (s, 2H, OCH₂O), 5.39 (s, 2H, NCH₂O), 6.49 (s, 2H, NH₂), 7.80 (s, 1H, 8-H), 10.61 (s, 1H, NH₂). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 169.4 (COOEt), 156.8 (6-C), 153.9 (2-C), 151.5 (4-C), 137.7 (8-C), 116.5 (5-C), 71.9 (N-CH₂-O), 65.9

(O-CH₂CO), 60.4 (OCH₂CH₃), 13.9 (CH₃). IR (KBr) 1764, 1687, 1631, 1575 cm⁻¹. MS (ESI) *m/z* 268 (MH⁺). HRMS (ESI) calcd for C₁₀H₁₄N₅O₄ (MH⁺): 268.1022. Found: 268.1066. Anal. Calcd for C₁₀H₁₃N₅O₄•H₂O: C, 42.10; H, 5.30; N, 24.56. Found: C, 42.31; H, 5.49; N, 24.30.

[(2-Amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]acetic acid (3).

Ethyl ester (**7**) (2.58 g, 9 mmol) was heated for 10 min in boiling water. After being cooled to rt, the mixture was treated with 10% NaOH (9 mL, 22.5 mmol) for 4 h under stirring. The mixture was neutralized with MeSO₃H (2.28 g, 24 mmol). The resulting white solids were washed with water and filtered. The solid was recrystallized from 25%AcOH to give **3** (2.13 g, 99%). mp>300°C. *R*_f=0.56 (CHCl₃/MeOH=3:1). ¹H NMR (300 MHz, DMSO-*d*₆) δ 4.12 (s, 2H, OCH₂O), 5.38 (s, 2H, NCH₂O), 6.51 (s, 2H, NH₂), 7.82 (s, 1H, 8-*H*), 10.61 (s, 1H, NH), 12.75 (s, 1H, COOH). ¹³C NMR (75.5 MHz) δ 168.1 (CO₂H), 153.9 (4-C), 151.1 (6-C), 148.6 (8-C), 134.9 (2-C), 113.7 (5-C), 69.1 (NCH₂O), 62.9 (OCH₂CO). IR (KBr) 1727, 1648, 1602 cm⁻¹. MS (ESI) *m/z* 240 (MH⁺). HRMS (ESI) calcd for C₈H₁₀N₅O₅ (MH⁺): 240.0733. Found: 240.0735. Anal. Calcd for C₈H₉N₅O₄: C, 40.17; H, 3.79; N, 29.29. Found: C, 40.08; H, 3.92; N, 28.97.

REFERENCES

1. D. M. Richards, A. A. Carmine, E. N. Brogden, R. C. Heel, T. M. Speight, and G. S. Avery, *Drugs*, 1983, **26**, 378.
2. J. J. O'Brien and D. M. Campoli-Richards, *Drugs*, 1989, **37**, 233.
3. R. J. Whitley and J. W. Gnann, Jr., *N. Engl. J. Med.*, 1992, **327**, 782.
4. M. J. Robins, P. W. Hatfield, J. Balzarini, and E. Declercq, *J. Med. Chem.*, 1984, **27**, 1486.
5. H. J. Schaeffer, L. Beauchamp, P. de Miranda, G. B. Elion, D. J. Bauer, and P. Collins, *Nature*, 1978, **272**, 583.
6. B. Lehmann, DE 19604101A (1997).
7. S. J. Culp, B. P. Cho, F. F. Kadlubar, and F. E. Evans, *Chem. Res. Toxicol.*, 1989, **2**, 416.
8. I. L. Shchaeleva, S. V. Kochetkova, T. L. Tsilevich, G. V. Vladyko, L. V. Korochenko, E. I. Boreko, I. P. Smirnov, A. A. Khorlin, B. P. Gottikh, and V. L. Florent'ev, *Bioorg. Khim. (Eng. Trans.)*, 1990, **15**, 523.